

1-Methylcyclopropene maintains postharvest quality in Norwegian apple fruit

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Abstract

Norwegian fruit production is mostly destined for the local market and can suffer from poor-quality retention during storage. 1-Methylcyclopropene (1-MCP) is an inhibitor of ethylene perception used to maintain the physical and functional quality of pome fruit. Extensive work has been carried out on the effect of 1-MCP on apples, but not on cultivars grown in Norway. In this work, the potential of 1-MCP application (0.625 ml l⁻¹ for 24 h at 0 ± 1°C) for ripening control of the apple cultivars ‘Aroma’, ‘Red Gravenstein’, and ‘Summered’ was studied during 1 and 1.5 months of cold storage; both scenarios were followed by five days of shelf life. The application of 1-MCP reduced softening by an average of 12% in ‘Aroma’, ‘Red Gravenstein’, and ‘Summered’ apples when cold stored for both 1 and 1.5 months as compared to control. External colour remained similar to initial values in 1-MCP fruit when compared to control apples, which presented a significant skin darkening. This indicated a delay in the ripening process. 1-MCP treatment did not affect total soluble solids content. ‘Aroma’ samples treated with 1-MCP showed a low sucrose hydrolysis, indicating a slower ripening process. This work confirms that 1-MCP postharvest treatment shows great potential for maintenance of apple cvs. in Norway during cold storage and shelf life.

Keywords *Malus domestica*, food waste, postharvest, quality

INTRODUCTION

Apples (*Malus domestica* Borkh) cultivated in Norway have a high initial quality and great market potential. They are highly appreciated by local consumers. The mean annual apple production in Norway (1998–2017) is only 12,660 t (FAOSTAT, 2018) which is mainly destined for the local market. These apples have poor quality retention during storage, with high index of firmness loss and fungal decay incidence.

Postharvest treatments can be applied to enhance local production and its availability. The application of controlled atmosphere (CA) storage is well established for apples, showing beneficial effects on quality retention by delaying ripening and senescence (Brizzolara et al., 2017; Falagán and Terry, 2018; Meberg et al., 2000; Thewes et al., 2017). Apples can be stored up to 12 months under near hypoxic conditions (Watkins, 2003), allowing year-round availability. In Norway, the targeted storage time is 1–2 months as this country’s local production is exclusively destined for seasonal consumption. For this reason, the investment in CA storage facilities is not cost-effective when compared to normal cold storage (Kart and Demircan, 2015).

Another option to delay ripening is controlling ethylene (Golding and Singh, 2017). 1-Methylcyclopropene (1-MCP) is a gaseous cyclic olefin which binds to ethylene receptors, avoiding ethylene-dependent responses (Almeida et al., 2016; Liguori et al., 2017; Sisler and Serek, 1997). Its effectiveness is extensively proven in many apples cvs. but not in apples grown in Norway, which could benefit from the ripening control action of 1-MCP (Gwanpua et al., 2017; Lee et al., 2016; Watkins and Nock, 2012; Watkins et al., 2000). 1-MCP was approved by the US Environmental Protection Agency for ornamental products in 1999 and for edible food products in 2002. Then, it was registered as Generally Recognised As Safe

active ingredient by the FDA (2004). By 2011, around 50 countries had approved the use of 1-MCP for fresh produce and there are currently five registered products that contain this active ingredient such as SmartFresh™ and EthylBlock™ by AgroFresh Inc. (Rademacher, 2015). In the European Union, the active ingredient 1-MCP is included in Annex I of the Directive 91/414/EEC (2005), allowing its use on fruits including apples. 1-MCP is a good alternative or supplement to CA storage because it can maintain fruit physical and functional quality across the supply chain (Blankenship and Dole, 2003; Golding and Singh, 2017; Watkins and Nock, 2012). The application of 1-MCP is straight forward, innocuous for the operator and facilities, and economical (Watkins, 2006). It should be applied along with cold storage practices, and preferably right after harvest. Its effectiveness on apple is mainly influenced both by the maturity stage of the fruit at harvest, and cultivar (cv.; Watkins, 2006). The main benefit of 1-MCP for apples is the reduction of major physiological storage disorders (Watkins et al., 2000) and firmness loss (Moran and McManus, 2005).

The aim of the work was to examine the potential of 1-MCP for control of ripening in the apple cvs.: ‘Aroma’, ‘Red Gravenstein’, and ‘Summered’, cultivated in Norway.

MATERIALS AND METHODS

Plant material

Apple cvs. ‘Aroma’, ‘Red Gravenstein’, and ‘Summered’ were harvested at commercial maturity from Sletthagen (5778 Utne, Norway) on 15 October 2018. Maturity was assessed by experienced local farm staff based on fruit appearance (i.e. size, skin colour, firmness). The samples were shipped by air on 16 October 2018, from Oslo (Norway) to the Plant Science Laboratory at Cranfield University (UK) under cold conditions ($\sim 0^{\circ}\text{C}$). The apples were immediately placed in the treatment boxes in a cold room at $0 \pm 1^{\circ}\text{C}$ and $90 \pm 5\%$ relative humidity (RH).

Experimental design

Apples of each cv. were placed in 300 l hermetically sealed boxes equipped with an inlet and outlet bulkhead fitting connected to an air supply system which recirculated the atmosphere inside the container. Two groups were considered: control (non-treated) and 1-MCP (AgroFresh™, UK), exposed to 0.625 ml l^{-1} for 24 h at $0 \pm 1^{\circ}\text{C}$ (Chope et al., 2007). The apples were stored at $0 \pm 1^{\circ}\text{C}$ and $90 \pm 5\%$ RH for one month followed by five days at 20°C simulating shelf life (Scenario 1), and 1.5 months at $0 \pm 1^{\circ}\text{C}$ and $90 \pm 5\%$ RH followed by five days at 20°C simulating shelf life (Scenario 2). Three biological replicates, consisting of six apples each, were analysed at each sampling point: day 0 – baseline; day 30 – after one month of cold storage; day 35 – after Scenario 1 shelf life (shelf life 1 – SL1); day 45 – after 1.5 months of cold storage; and day 50 – after Scenario 2 shelf life (shelf life 2 – SL2). Samples from each cv. were snap-frozen and freeze dried at each evaluation date. Samples were kept at -80°C until analysis.

Firmness

Firmness was measured with a uniaxial testing machine (Instron 5542, Instron, Bucks., UK). A 10 mm diameter probe was used at 240 mm min⁻¹ crosshead speed and 8 mm penetration depth, according to Anastasiadi et al. (2017). Results were expressed as Newton (N).

Colour

For each replicate, six apples were placed in a photoE-Box plus 1419 (Ortech Professional Lighting, CA, USA). White fluorescent light (D65) was used on the sides and underneath the fruit. An exposure of 62 ms, intensity of 155, gain of 0.9, and gamma of 1 were applied. Absorption spectrum was recorded and analysed using a Lumenera Infinity3-6UR camera

(Lumenera corporation, Ottawa, Canada) and the Infinity Analyse application package. Although measurements of L^* , a^* , and b^* ; chroma; and hue angle (H°) were recorded, we herein reported H° as it best represents the colour change between green and red ($H^\circ = \frac{1}{4} \tan^{-1} [b^*/a^*]$; Duarte et al., 2009).

Total soluble solids

Total soluble solids were determined using a digital refractometer (model PR-32a, Atago Ltd, Tokyo, Japan). Results were expressed as $^\circ$ Brix.

Non-structural carbohydrates

For each replicate, freeze-dried apple powder (150 mg) was extracted according to Downes and Terry (2010) with slight modifications. Skin and flesh were analysed separately. An HPLC (Agilent Technologies 1200 series, Berks., UK) with an evaporative light scattering detector (Agilent Technologies 1200 Series, G1362A) was used to quantify sugar extracts (20 ml). Elution was performed at 1 ml min⁻¹ flow rate, at 30°C, as follows: 80% B at 0 min, 50% at 15 min, 50% at 18 min, and 80% at 20 min. Sugars were quantified using the external standards: glucose, fructose, and sucrose purchased from Sigma-Aldrich.

Statistical analysis

Data were subjected to analysis of variance using GenStat for Windows (8.1, VSN International Ltd, Herts., UK). The differences between cvs., treatments, and storage time were studied. Least significant difference (LSD) values ($p < 0.05$) were calculated from each analysis.

RESULTS AND DISCUSSION

Meeting consumer demands is key to introducing new cvs. in the market (Silvestri et al., 2018). In the case of apples, it is the taste (i.e. sweetness, texture, mouth-feel attributes) and/or appearance (i.e. external fruit colour, defects), which drives consumer choice (Chen et al., 2017; Pre-Aymard et al., 2005). The marketability of the three main apple cvs. grown in Norway quickly declines after harvest, as they lose their firmness, negatively impacting on their quality. This process critically reduces the consumer acceptability.

Investing in CA facilities is not ideal as, in this case, the aimed storage period is short (1–2 months). In this study, 1-MCP treatment was applied to maintain the initial quality of apples cultivated in Norway by delaying ripening. Apple firmness is broadly used as a ripening and ‘condition’ indicator. The consumer considers that around 50 N is the best eating firmness level for apple fruit (Hoehn et al., 2003). ‘Red Gravenstein’ apples showed the highest initial firmness (~45 N) when compared to ‘Aroma’ and ‘Summered’ cvs. (~35 N). In all cases the values were lower than the firmness at harvest of other well-known cvs. such as ‘Royal Gala’ which is generally picked at 65–70 N (Schwallier, 2012). For this reason, maintaining the values registered at harvest is fundamental to stay close to the 50 N consumers’ requirements. The firmness values at day 0 were better maintained in fruit subjected to 1-MCP treatment compared to control fruit in both scenarios. Specifically, ‘Aroma’ apples treated with 1-MCP showed no loss in firmness after 1 or 1.5 months of cold storage, while control ‘Aroma’ apples lost ca. 10% of their initial firmness after one month of cold storage and 9.22% after 1.5 months. Similar results were obtained for ‘Red Gravenstein’ and ‘Summered’ cvs. (Figures 1 and 2). Apple softening is related to the disassembly of the cell wall components (pectin and hemicellulose) that affects turgor and fruit water status (Atkinson et al., 2012). This process is regulated by ethylene, activating softening-associated enzymes such as polygalacturonase (Rupasinghe et al., 2000).

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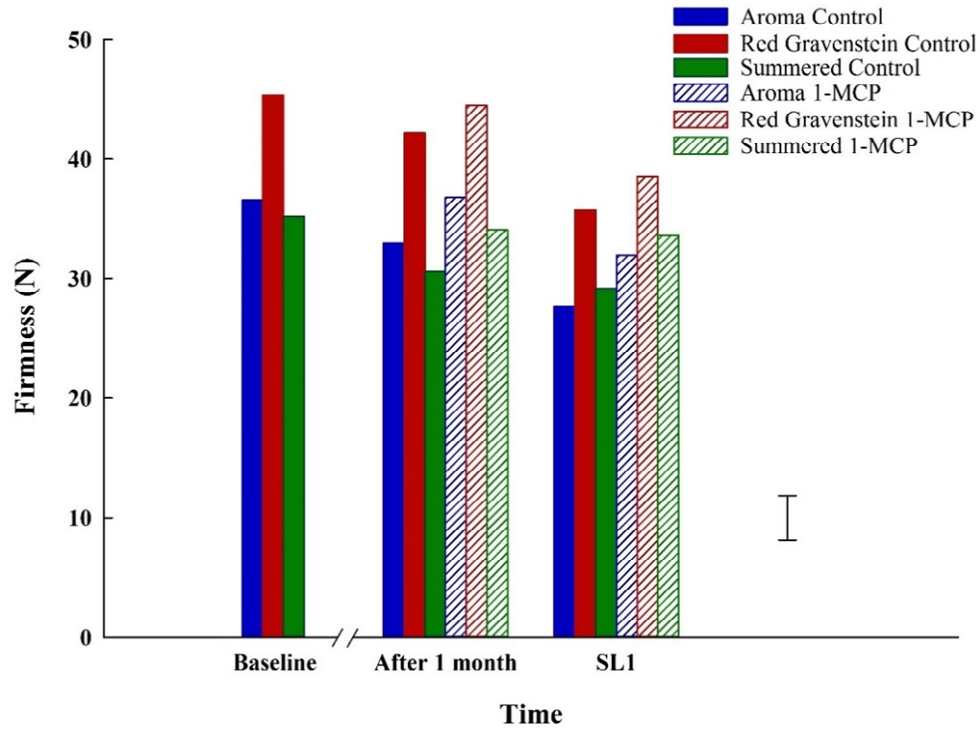


Figure 1. Firmness (N) of 'Aroma', 'Red Gravenstein' and 'Summered' apples subjected to Control (non-treated) and 1-MCP ($0.625 \mu\text{L L}^{-1}$, 24 h, 0°C) is shown at day 0 (Baseline), after 1 month of cold storage (After 1 month), and after 5 days of room temperature storage, simulating home shelf life (SL1). Bars represent means of three replicates ($n = 10$). $\text{LSD}_{\text{treatment} \times \text{time point}}$ is shown.

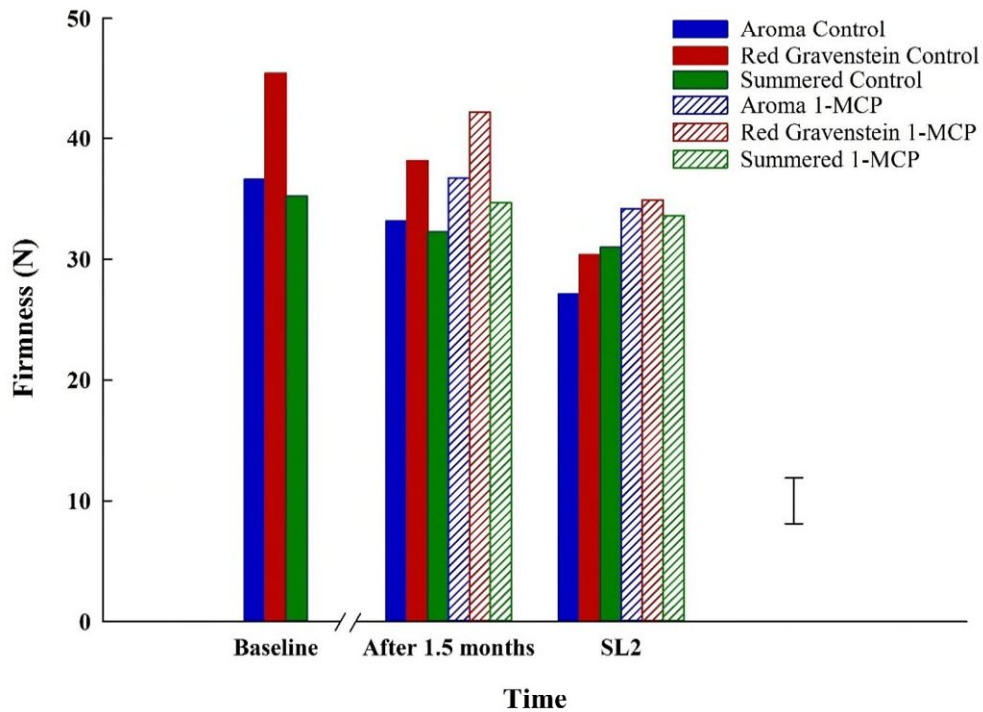


Figure 2. Firmness (N) of 'Aroma', 'Red Gravenstein' and 'Summered' apples subjected to Control (non-treated) and 1-MCP ($0.625 \mu\text{L L}^{-1}$, 24 h, 0°C) is shown at day 0 (Baseline), after 1.5 months of cold storage (After 1.5 months), and after 5 days of room temperature storage, simulating home shelf life (SL2). Bars represent means of three replicates ($n = 10$). $\text{LSD}_{\text{treatment} \times \text{time point}}$ is shown.

1-MCP binds to ethylene receptors, suppressing ethylene-related responses. The interaction of 1-MCP with the apple ethylene receptors would likely have prevented loss of firmness in 1-MCP treated fruit compared to control apples, which lost an average of 10% after one month of cold storage and 12.85% after 1.5 months.

Temperature is an essential factor determining fresh produce postharvest quality (Paull, 1999). However, fruit is often exposed to non-optimal temperatures during transport, distribution, retail, and household storage (Johnston et al., 2001), severely affecting quality. This can result in rejections, impacting on food waste. In the case of apples, 0–3°C is the optimal temperature range, depending on the cv. In order to simulate retailer and shelf life, apples were exposed to room temperature (~20–25°C) after cold storage. We speculated that this increase in temperature stimulated greater ethylene production (Gwanpua et al., 2017), which meant a reduction of firmness (Figures 1 and 2). The temperature impact was greater in control fruit than 1-MCP apples as control lost 12.46 and 16% more firmness than 1-MCP fruit in SL1 (1 month) and SL2 (1.5 months), respectively. With respect to the two supply chain scenarios considered, ‘Aroma’ and ‘Red Gravenstein’ apples had a better firmness score when stored for 1 month, while ‘Summered’ fruit had a better response when stored for 1.5 months (Figures 1 and 2).

Colour is also an important parameter for decision-making when it comes to choosing a variety: it is classified as a ‘search attribute’ together with price. Background colour change from green to yellow is a sign of an active ripening process and therefore, indicating a loss of freshness. The background yellow/green colour of the apple peel is due to chlorophyll and carotenoids; the apple red colour is produced by anthocyanins and flavonols (Lancaster and Dougall, 1992). Anthocyanin biosynthesis is regulated by ethylene and external factors such as sun exposure, accumulating in the skin tissue during the ripening process (Dar et al., 2019). Characteristically, ‘Red Gravenstein’ apples were redder than ‘Aroma’ and ‘Summered’ cvs. on day 0, showing a significantly lower H° . After one month of cold storage, fruit treated to 1-MCP treatment did not change their initial colour, while control conditions reduced H° level in 23.53, 9.70, and 41.12% for ‘Aroma’, ‘Red Gravenstein’, and ‘Summered’ apples, respectively. This meant an increase in skin reddening, reducing the fresh appearance of the fruit and was consistent with other studies where the loss of greenness of the background colour was inhibited by 1-MCP (Saftner et al., 2003; Zanella, 2003). After 1.5 months of cold storage, control fruit reduced their H° values on average 31%, which implied a darkening in the skin colour (Figure 4), while no significant differences were shown by 1-MCP treated apples. The increase of temperature in the shelf life period affected colour development. All fruit presented a decrease of the H° values, especially those stored under control conditions (Figures 3 and 4). This result agreed with other studies which showed that storage temperature was involved in triggering senescence and ripening during storage (Lee et al., 2016; Mir et al., 2001).

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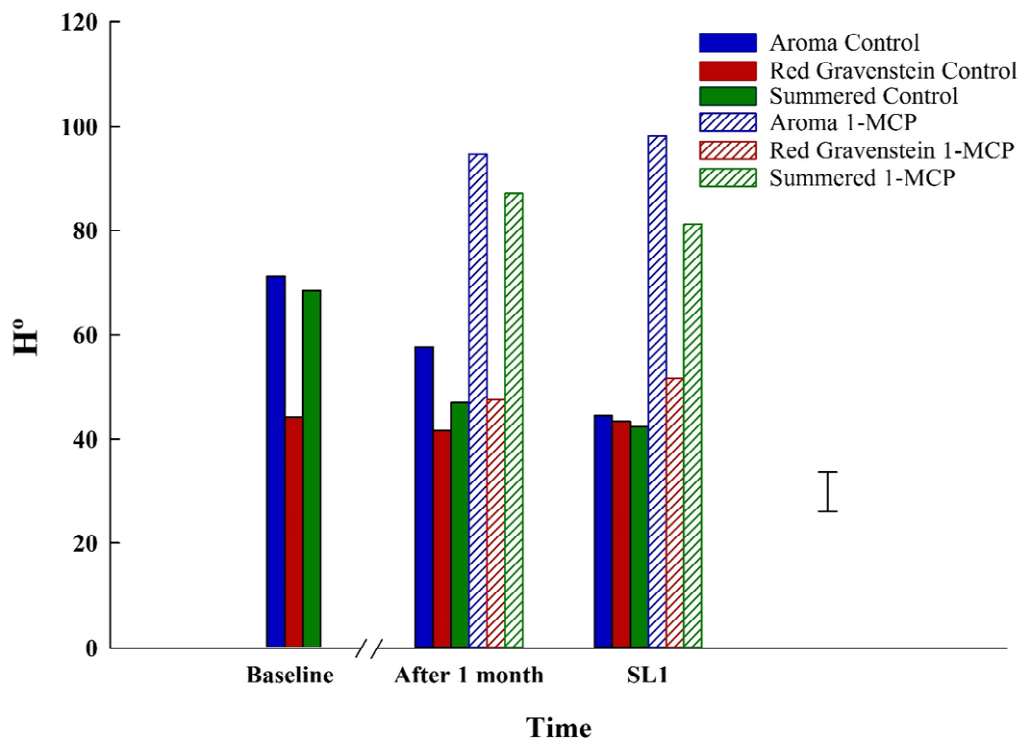


Figure 3. Hue angle (H°) of ‘Aroma’, ‘Red Gravenstein’ and ‘Summered’ apples subjected to Control (non-treated) and 1-MCP ($0.625 \mu\text{L L}^{-1}$, 24 h, 0°C) is shown at day 0 (Baseline), after 1 month of cold storage (After 1 month), and after 5 days of room temperature storage, simulating home shelf life (SL1). Bars represent means of three replicates ($n = 10$). $\text{LSD}_{\text{treatment} \times \text{cv}}$ is shown.

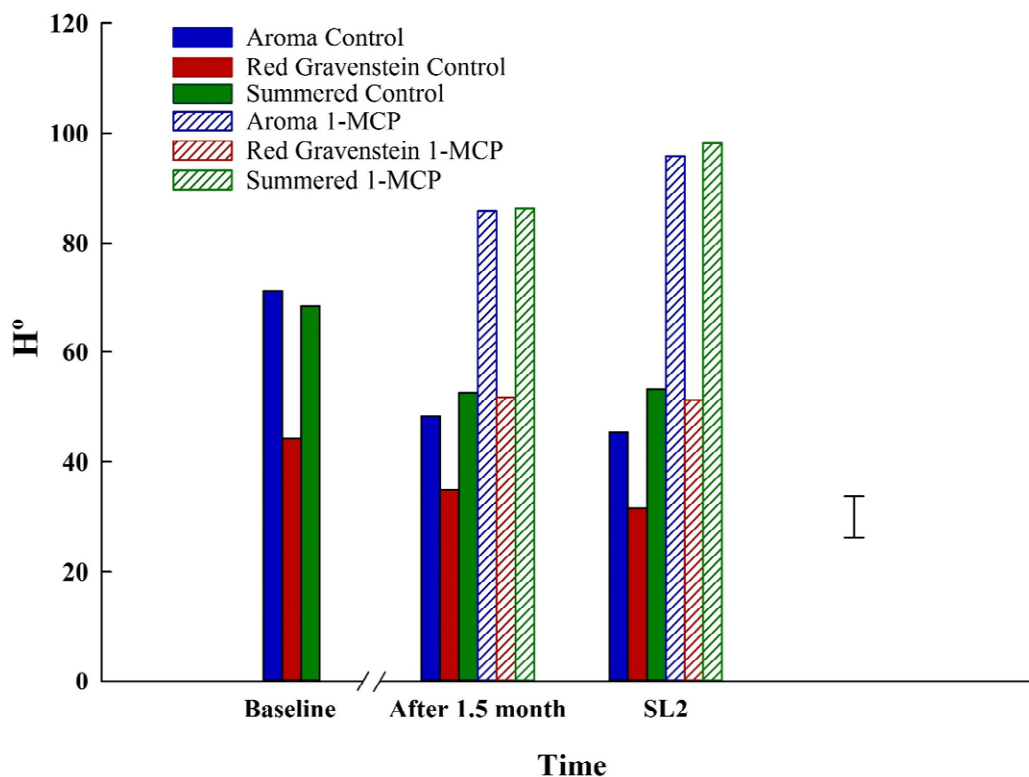


Figure 4. Hue angle (H°) of ‘Aroma’, ‘Red Gravenstein’ and ‘Summered’ apples subjected to Control (non-treated) and 1-MCP ($0.625 \mu\text{L L}^{-1}$, 24 h, 0°C) is shown at day 0 (Baseline), after 1 month of cold storage (After 1.5 months), and after 5 days of room temperature storage, simulating home shelf life (SL2). Bars represent means of three replicates ($n = 10$). $\text{LSD}_{\text{treatment} \times \text{cv}}$ is shown.

Consumers choose an apple cv. based on its firmness and appearance, but their decision to repurchase depends on their satisfaction to its organoleptic characteristics (Harker et al., 2003). Sweetness is one of the drivers for consumer preference and TSS is used to estimate this trait (Aprea et al., 2017). The accumulation of TSS during fruit development determines sweetness (Li et al., 2018) and is used as an indicator for decision-making on selecting an appropriate picking date. In this work, ‘Summered’ apples showed the highest TSS at harvest when compared to ‘Aroma’ and ‘Red Gravenstein’ cvs. ($13.32 \pm 0.15^\circ\text{Brix}$ vs. 11.12 ± 0.19 and $12.52 \pm 0.20^\circ\text{Brix}$, respectively). TSS content generally changes with fruit development, peaking at maturity (Desnoues et al., 2014). However, no differences were found after 1 and 1.5 months of cold storage with respect to the initial values, for both control and 1-MCP treatments (data not shown).

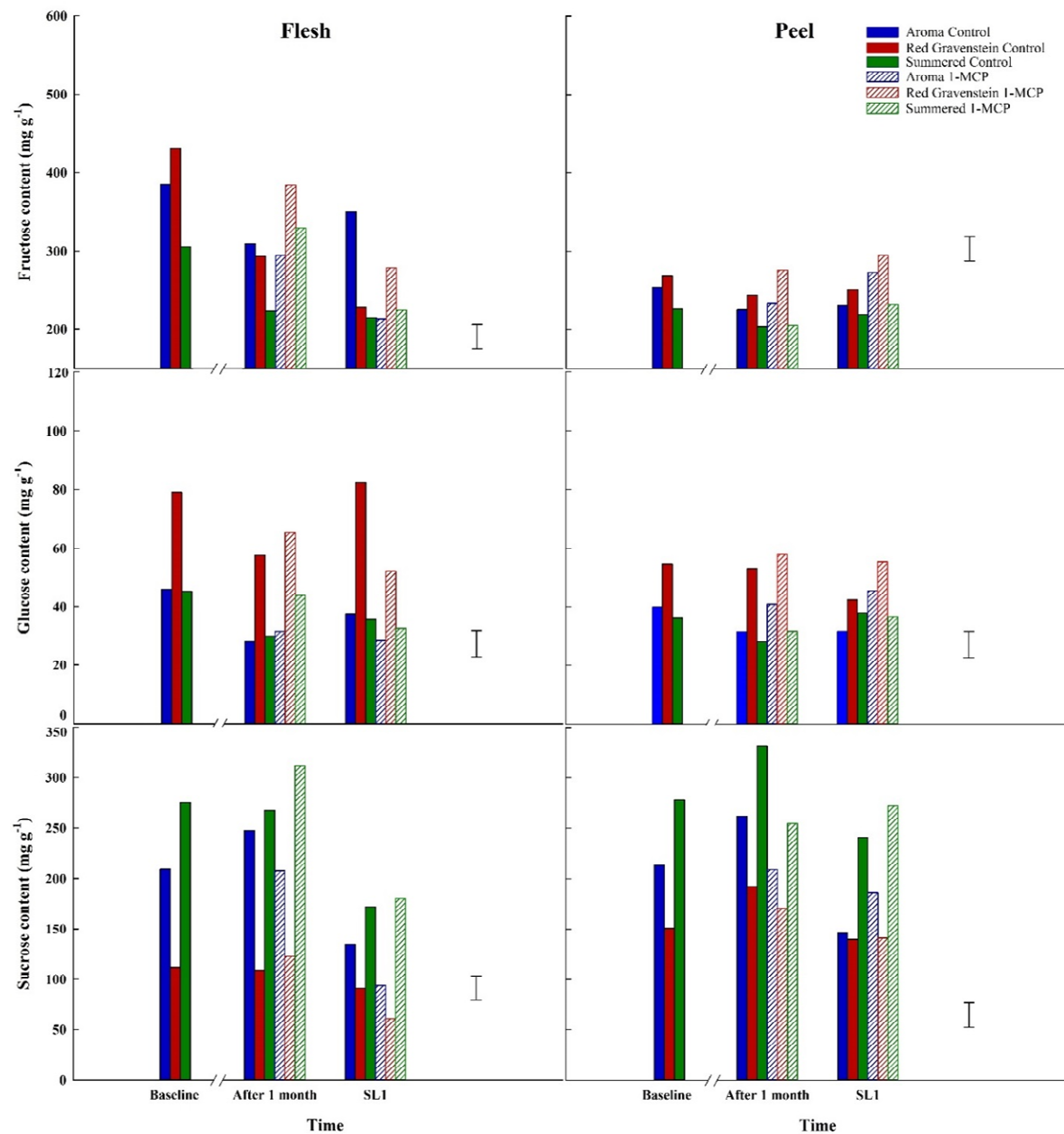


Figure 5. Reducing sugars (Fructose, Glucose, Sucrose; mg g⁻¹) of ‘Aroma’, ‘Red Gravenstein’ and ‘Summered’ apples subjected to Control (non-treated) and 1-MCP (0.625 μL L⁻¹, 24 h, 0 °C) is shown at day 0 (Baseline), after 1 month of cold storage (After 1 month), and after 5 days of room temperature storage, simulating home shelf life (SL2). Bars represent means of three replicates (n = 3). LSD_{time point * cv} is shown.

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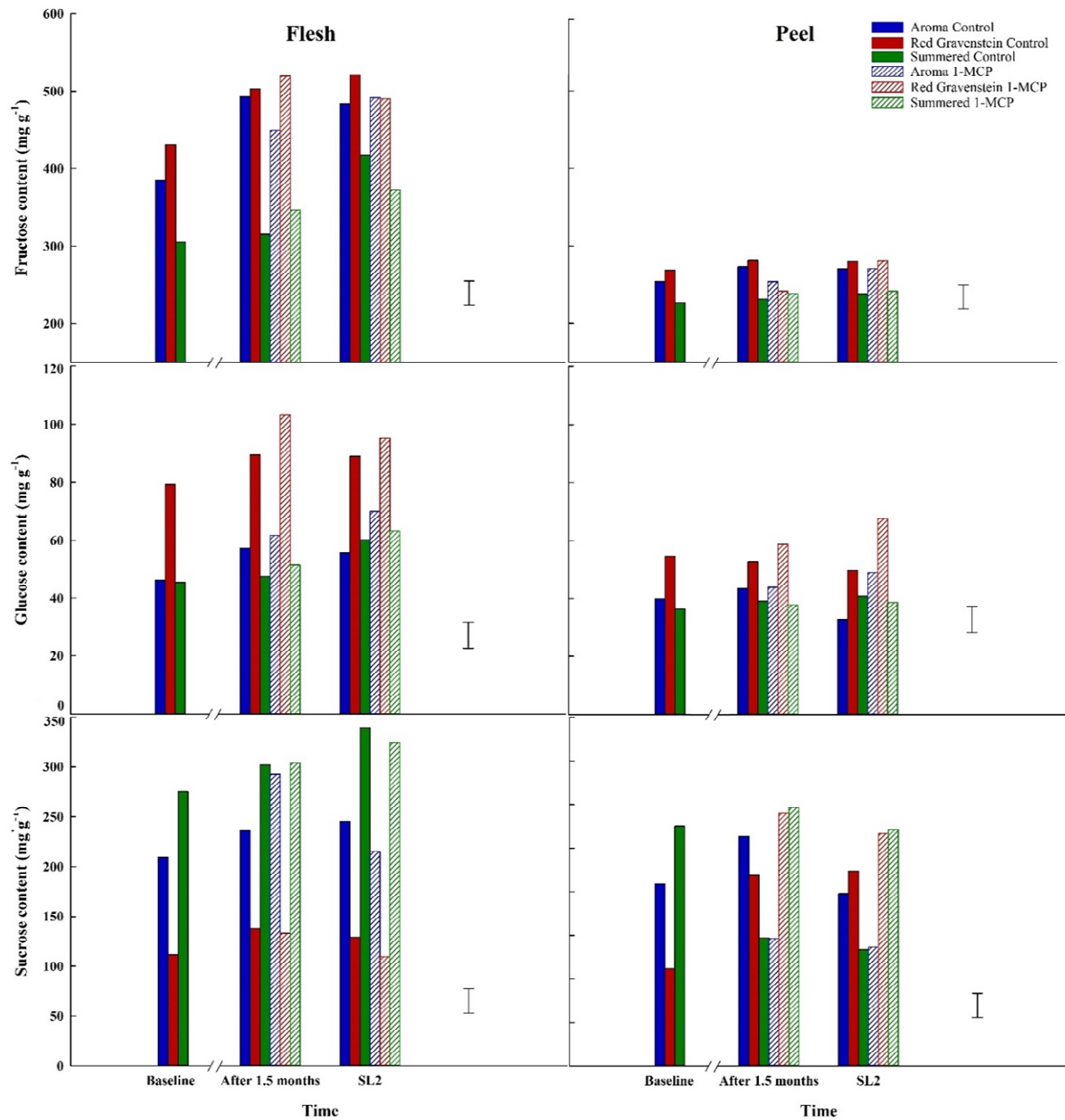


Figure 6. Reducing sugars (Fructose, Glucose, Sucrose; mg g^{-1}) of ‘Aroma’, ‘Red Gravenstein’ and ‘Summered’ apples subjected to Control (non-treated) and 1-MCP ($0.625 \mu\text{L L}^{-1}$, 24 h, 0°C) is shown at day 0 (Baseline), after 1.5 months of cold storage (After 1.5 months), and after 5 days of room temperature storage, simulating home shelf life (SL2). Bars represent means of three replicates ($n = 3$). $\text{LSD}_{\text{time point} \times \text{cv}}$ is shown.

It has been reported that 1-MCP can increase, decrease, or not affect the TSS depending on the apple cv. (Watkins, 2000), which was also observed in this study. We hypothesised that an earlier harvest date could increase the efficacy of 1-MCP, as the number of ethylene receptors would be lower and therefore, easier to control with a 1-MCP treatment (Tatsuki et al., 2007). In the case of non-structural carbohydrates, fructose was the main sugar for all cvs. (Figures 5 and 6), in agreement with Li et al. (2012). ‘Red Gravenstein’ showed the highest fructose content at harvest when compared to ‘Aroma’ and ‘Summered’ cvs. This result indicated that ‘Red Gravenstein’ apples were sweeter than ‘Aroma’ and ‘Summered’ as fructose directly impacts on sweetness perception, being almost twice as sweet as glucose (Biester et al., 1925;

Desnoues et al., 2014). Fructose accumulated more in the flesh than skin. In both scenarios, both control and 1-MCP apples showed a slight decrease in fructose concentration, with a greater increase under control conditions (Figures 5 and 6).

In apples, a small portion of fructose is converted into starch; in general, it accumulates in the vacuoles of the tissue cells with maturation (Aprea et al., 2017). Correspondingly, fructose content is higher than glucose content as confirmed by this work. Glucose is generally incorporated into starch, showing lower concentrations in fruit tissue (Yamaki and Ino, 1992). At harvest, 'Red Gravenstein' apples had the highest glucose concentration when compared to 'Aroma' and 'Summered' cvs. (for flesh: 79.30 ± 2.21 mg g⁻¹ dw vs. 46.27 ± 0.77 and 45.62 ± 0.22 mg g⁻¹ dw, respectively; for skin: 54.82 ± 1.12 mg g⁻¹ dw vs. 39.87 ± 0.17 and 36.30 ± 0.77 mg g⁻¹ dw, respectively). All samples presented a slight decrease during storage and shelf life (Figures 5 and 6). In the case of sucrose, 'Summered' apples showed the highest content when compared to 'Aroma' and 'Red Gravenstein' cvs. (for flesh: 275.20 ± 4.84 mg g⁻¹ dw vs. 209.20 ± 1.09 and 111.70 ± 7.88 mg g⁻¹ dw, respectively; for skin: 278 ± 3.27 mg g⁻¹ dw vs. 213.30 ± 1.64 and 150.81 ± 3.27 mg g⁻¹ dw, respectively). Sucrose content showed a subtle decreasing trend in flesh in scenario 1 while maintaining the initial values in scenario 2 (Figures 5 and 6). According to the behaviour patterns of individual sugars with respect to storage and shelf life time, sucrose decreased for most lines, while fructose increased for 'Aroma' control samples, confirming the theory of sucrose hydrolysis. During postharvest period, sucrose suffers an irreversible hydrolysis into fructose and glucose, catalysed by the enzyme invertase (Kleczkowski et al., 2010; Tong et al., 2018). 1-MCP treatment had no significant effect on glucose and sucrose contents (Figures 5 and 6). However, fruit exposed to 1-MCP showed a lower increase in fructose content ($p < 0.05$). This indicated a slower ripening as sugar metabolism is linked to plant processes, regulating plant development (Li et al., 2012). In this sense, the effect of 1-MCP on ripening was observed in both the quality results and the biochemical compounds.

The ratios between the different individual sugars are also involved in the apple fruit taste; in particular, the ratio fructose-to-glucose (Desnoues et al., 2014). 'Red Gravenstein' showed the highest fructose and glucose concentrations in both peel and flesh; it was the cv. which showed the lowest fructose-to-glucose ratio when compared to 'Aroma' and 'Summered' cvs. (for flesh: 7.63 vs. 11.59 and 10.18, respectively; for skin: 7.10 vs. 9.24 and 9.74, respectively). These results positively correlated with the TSS measurements. Fructose-to-glucose ratio decreased over time in all cases, especially during shelf life. For example, 'Aroma' apples stored under control conditions reduced their fructose-to-sucrose ratio by 9% after cold storage, while during shelf life the ratio decreased 22.70% with respect to the initial values. Despite the decrease, the cvs. had a much higher fructose-to-glucose ratio than the apple cvs. currently available in many supermarkets (Granny Smith: 2.1; Pink Lady: 3.4; Golden Delicious: 2.6; Royal Gala: 3.0; Fuji: 2.3; Hermann and Bordewick-Dell, 2018).

CONCLUSION

The data shown in this work has confirmed that 1-MCP postharvest treatment has a promising potential for maintenance of apple cvs. grown in Norway during cold storage. Its application is a good alternative to CA methods. Further research is needed to test if an earlier harvest could improve the efficacy of 1-MCP in apples grown in Norway.

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